

Clinical Involvement of the Tonsillar Immune System in IgA Nephropathy

MARIE C. BÉNÉ¹, GILBERT C. FAURE¹, BRUNO HURAUULT DE LIGNY² and ANNE KENNEL DE MARCH¹

From the ¹Laboratoire d'Immunologie du CHU, Faculté de Médecine de Nancy, 54500, Vandoeuvre les Nancy, France, and ²Service de Néphrologie, CHU de Caen, 14000 Caen, France

Béné MC, Faure GC, Hurault de Ligny B, Kennel de March A. *Clinical involvement of the tonsillar immune system in IgA nephropathy.* Acta Otolaryngol 2004; Suppl 555: 10–14.

The temporal association of tonsillitis and hematuria or proteinuria in IgA nephropathy suggests that there might be a link between the physiological properties of the secondary lymphoid organ that tonsils represent and the mesangial deposition of IgA characteristic of this nephropathy. A number of clinical and *ex-vivo* data support this hypothesis. One of the earliest was the demonstration of the dimeric nature of mesangial IgA, composed of IgA monomers linked by a J chain, yet lacking the polyIg receptor acquired by secretory IgA during transcytosis through epithelial cells. This molecular structure is that of IgA synthesized in human tonsils, the epithelium of which lacks polyIg receptor. Moreover, tonsils from patients with IgA nephropathy display an abnormal partition of IgG and IgA producing plasma cells associated with a significantly developed web of high endothelial venules. IgA nephropathy could thus be in part related to an alteration of IgA precursors homing in tonsils. Tonsillectomy thus would present the advantage of removing an abnormally functioning source of dimeric IgA. Performed early enough in the course of the renal disease, tonsillectomy could suffice to halt the development of the nephropathy and restore the kidneys to health. *Key words:* dimeric IgA secretion, human tonsils, mesangial deposits.

INTRODUCTION

IgA Nephropathy (IgAN), as defined in 1968 by Berger and Hinglais (1) is characterized by the deposition of IgA in glomerular mesangium. This mesangial anomaly progressively leads to glomerular sclerosis, ultimately resulting in kidney failure. Several reports from the literature suggest that IgA are transiently deposited in the kidney, in a more or less continuous fashion. This is supported by the fact that IgAN patients often display bouts of hematuria or proteinuria, suggesting remitting/relapsing mesangial alterations. Moreover, it has been reported that transplanted kidneys from donors with previously undetected IgAN could “heal”, i.e. clear the IgA deposits. Conversely, IgAN frequently relapses after transplantation, at least through the appearance of IgA deposits in the transplant (2, 3). These data suggest that mesangial IgA in IgAN originate from an extra renal source.

IGA SOURCES IN THE BODY

IgA represent the second most abundant isotype of immunoglobulins in plasma, after IgG. They also are the predominant isotype in secretions. Plasmatic and secretory IgA differ in their molecular form. The former are monomers, mostly of IgA1 subclass, while secretory IgA are synthesized as dimers, linked by a J chain (4). Secretory IgA reach the lumen of mucosal spaces by crossing the lining of the epithelium through the active use of the epithelial PolyIg receptor (PigR) which is cleaved from the apical pole of epithelial cells and remains attached as protection to

secretory IgA. IgA2 are also more common among secretory IgA.

Plasmatic IgA have been widely studied in IgAN patients. It is thus well established that such patients often have increased serum IgA levels (5), with increased high molecular weight IgA (6). These IgA are polyclonal and no restriction pattern can be seen in high resolution electrophoresis, although the pI range of IgA from IgAN patients tends to be more restricted than in controls (7). This could be due to the glycosylation anomalies reported in IgAN by several groups (8, 9). More precisely, an abnormal partition of galactosyl residues on IgA carbohydrate side chains has been reported (10, 11) that could be related to a defect in the enzymatic system of galactosylation. However, no anomaly of the galactosyl synthase genes has been observed so far (12).

The various physiological IgA sources have also been studied in IgAN. Anomalies have been reported in the bone marrow (13, 14) and in tonsils (15–20) while normal data were obtained from the study of gut samples (21, 22). There are no available data from the lymph nodes or spleen.

The molecular form of IgA in the mesangium has also been a topic of interest for several groups. IgA1 were found to be nearly exclusive (13) while both IgA1 and IgA2 were observed by our group (23) and others (24, 25). Mesangial IgA in IgAN also were demonstrated to contain J chain yet lack PigR (23), a molecular structure compatible with that of mucosal IgA before transcytosis, which was one of the major conceptual breakthroughs initiating the mucosal hypothesis of IgA nephropathy.

THE MUCOSAL HYPOTHESIS

According to the data reported above, a pathophysiological hypothesis of IgAN could be that dimeric IgA are abnormally produced in a lymphoid organ, reach the peripheral blood, and, because of their size (and possibly charge) fail to be properly filtered by the kidneys but rather accumulate in the mesangium.

The possibility that palatine tonsils (or perhaps the whole Waldeyer ring) could be that extra-renal source of IgA is supported by several observations and studies. Recurrent ENT infections are reported by more than 70% of IgAN patients, often since childhood. These patients also report what was dubbed "synpharyngitic microhematuria" by Clarkson et al. (26), i.e. bouts of micro- or even macro-hematuria within 48 h of the onset of tonsillitis. Close examination of palatine and pharyngeal tonsils often discloses an enlargement in IgAN patients, considered pathologic enough by many ENT surgeons to sustain tonsillectomy.

Physiologically, tonsils are important lymphoid organs of the mucosae-associated lymphoid tissue. They contain germinal centers and plasma cells but produce no IgR. The immunoglobulins they synthesize drain from tonsils via blood and lymph vessels, and thus probably constitute a good part of plasmatic Ig. As in the rest of the MALT, cell recirculation is a major phenomenon in human tonsils. This physiological idiosyncrasy of the MALT involves anatomical "inducer" sites, which are mostly represented by Peyers' patches and solitary nodules in the gut, and "effector" sites consisting of the diffuse lymphoid tissue lining all mucosae. Human tonsils could be an inductor site, but also behave as an effector site because of their richness in plasma cells. Lymphocyte recirculation implies that specific cells activated in inducer sites then leave these tissues, through lymphatic vessels, reach the peripheral blood, then relocate in lymphoid tissues or mucosal areas containing high endothelial venules (HEV). This allows a dissemination of exquisitely adapted specific immunity to all mucosal territories.

In physiological circumstances, human tonsils contain about 60% of IgG secreting plasma cells and 40% of IgA secreting plasma cells. However, in IgAN, these proportions are reversed, with 60% or more of IgA producing plasma cells (15, 16, 18, 19). This anomaly is associated with a highly developed web of HEV in IgAN patients, significantly more important than in controls. These HEV express high levels of adhesion molecules involved in lymphocyte recirculation (27).

The "tonsillar hypothesis" of IgAN pathophysiology would thus involve an activation of the mucosal immune system by environmental antigens, followed

by either a higher output of IgA producing precursors and/or an increased trapping of these cells by activated HEV in tonsils.

Indeed, the study of peripheral blood cells in IgAN (28) shows significantly lower levels of IgA-secreting cells detected in ELISA spot, which is consistent with increased trapping of these cells. Also in support of such anomalies is the demonstration of a modified expression of adhesion molecules on peripheral B-cells, with significantly higher percentages of CD62L+/CD31+ B-cells with a significantly higher density of these molecules on the cells' surface (29). These data are both in favour of an increased activation of the MALT by environmental antigens and a better trapping of these cells by HEV. Increased activation is also supported by the fact that tonsillar lymphocytes from IgAN patients spontaneously proliferate more than cells from controls, as demonstrated by the significantly higher proportions of cells in S and G2 phase when analysing the cell cycle of freshly eluted tonsillar cells.

It has been shown that IgA produced by tonsillar B lymphocytes from IgAN patients were able to bind to the mesangium of human kidney glomeruli (30). Germinal center B cells appear to contain more B-1 (CD5+) cells with reduced susceptibility to Fas-mediated apoptosis in IgA nephropathy tonsils (31). Regarding the antigenic specificity of the Ig response of B cells, a tendency towards respiratory environmental antigens has been observed by us (unpublished results). In Japan, it has been reported that *Haemophilus parainfluenzae* antigens were able to stimulate preferentially T and B cells from tonsils of IgAN patients (32).

TONSILLECTOMY IN IGAN

A logical conclusion of all the considerations depicted above would be that tonsillectomy, rather benign surgery, could benefit IgAN patients by removing the source of abnormal IgA.

Tonsillectomy has been proposed by several authors (33–35) but the demonstration of the efficacy of this procedure is suffering from the fact that it cannot easily comply to classical criteria of randomized double blind therapeutic evaluation. Recently, a methodologically-sound study was however published, supporting the interest of tonsillectomy in IgAN (36).

Indeed, our group still has regular news from the first patient in whom tonsillar IgA plasma cells predominance was seen by us in 1983: He had a definite diagnosis of IgAN confirmed by kidney biopsy immunofluorescence. Since his tonsillectomy, he has not had any episodes of proteinuria or

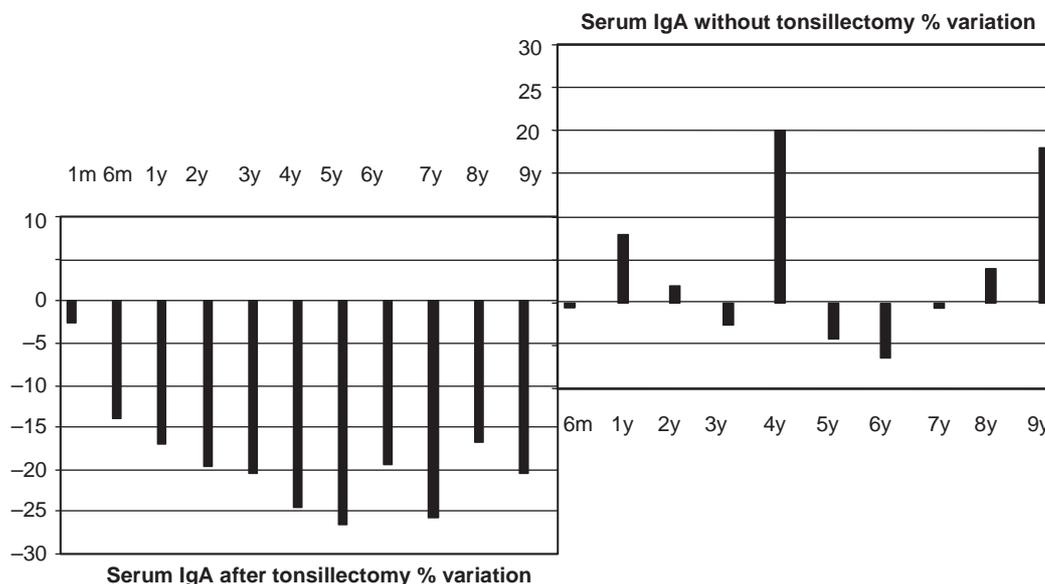


Fig. 1. Evolution over time of serum IgA levels in IgAN patients with or without tonsillectomy. Serum IgA were assayed repeatedly in 50 tonsillectomized IgAN patients and 21 patients who were not tonsillectomized. Follow-up was available for 9 years in both groups. A significant decrease in serum IgA was observed in tonsillectomized patients only, confirming tonsils as a possible source of abnormal IgA transiting towards kidney glomeruli via the peripheral blood.

hematuria and has not needed to see the nephrologist since.

In order to confirm this impression, we initiated a prospective series of IgAN patients follow-up (37). The cohort included 71 patients with identical renal parameters at inclusion, i.e. still mild glomerulonephritis. Among them, 50 were tonsillectomized and 21 were not. As shown in Fig. 1, plasmatic IgA levels rapidly and significantly decreased in tonsillectomized patients, while, overall, there was either no variation or an increase in non-tonsillectomized patients. Among tonsillectomized patients, 16 rapidly became lost to

follow-up, usually because they did not feel the need to consult a nephrologist any more. For the 34 patients available for follow-up, the highly satisfactory evolution of creatinemia and proteinuria is shown in Fig. 2.

CONCLUSION

A number of clinical and *ex-vivo* experimental indicators suggest that there is a rationale for considering therapeutic tonsillectomy in IgAN. However, it must be kept in mind that as tonsillectomy is merely removing the abnormal source of IgA, it will have no positive effect of the nephropathy if it is performed too late. Encouraging data has come from the recent Japanese studies. Tonsillectomy undertaken in children will, with time, prove most important as it might eventually demonstrate that IgAN incidence in this cohort was drastically reduced by this simple surgical procedure.

REFERENCES

- Berger J, Hinglais N. Intercapillary deposits of IgA-IgG. *J Urol Nephrol (Paris)* 1968; 74: 694-5.
- Berger J, Noel LH, Nabarra B. Recurrence of mesangial IgA nephropathy after renal transplantation. *Contrib Nephrol* 1984; 40: 195-7.
- Berger J. Recurrence of IgA nephropathy in renal allografts. *Am J Kidney Dis* 1988; 12: 371-2.

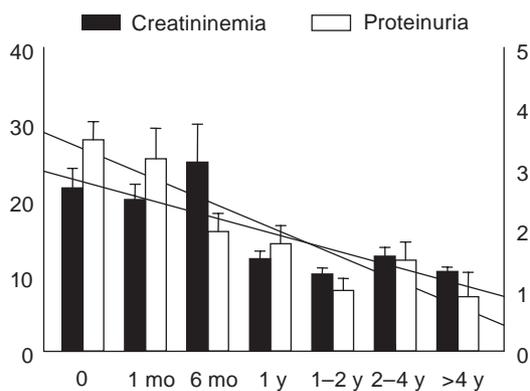


Fig. 2. Similarly, creatinemia and proteinuria were measured after tonsillectomy in 32 IgAN patients available for over 4 years of follow-up and a steady and significant decrease was observed over time. This suggests that tonsillectomy, by decreasing the amount of abnormal IgA, allows for kidney healing.

4. Johansen FE, Braathen R, Brandtzaeg P. Role of J chain in secretory immunoglobulin formation. *Scand J Immunol* 2000; 52: 240–8.
5. D'Amico G. Idiopathic IgA mesangial nephropathy. *Nephron* 1985; 41: 1–13.
6. Valentijn RM, Kauffmann RH, de la Riviere GB, Daha MR, Van ES LA. Presence of circulating macromolecular IgA in patients with hematuria due to primary IgA nephropathy. *Am J Med* 1983; 74: 375–81.
7. Harada T, Hobby P, Courteau M, Knight JF, Williams DG. Charge distribution of plasma IgA in IgA nephropathy. *Clin Exp Immunol* 1989; 77: 211–4.
8. Tomino Y, Ohmuro H, Takahashi Y, Suzuki Y, Saka S, Tashiro K, Shirato I, Koide H. Binding capacity of serum IgA to jacalin in patients with IgA nephropathy using jacalin-coated microplates. *Nephron* 1995; 70: 329–33.
9. Mestecky J, Hashim OH, Tomana M. Alterations in the IgA carbohydrate chains influence the cellular distribution of IgA1. *Contrib Nephrol* 1995; 111: 66–71.
10. Horie A, Hiki Y, Odani H, Yasuda Y, Takahashi M, Kato M, Iwase H, Kobayashi Y, Nakashima I, Maeda K. IgA1 molecules produced by tonsillar lymphocytes are under-O-glycosylated in IgA nephropathy. *Am J Kidney Dis* 2003; 42: 486–96.
11. Itoh A, Iwase H, Takatani T, Nakamura I, Hayashi M, Oba K, Hiki Y, Kobayashi Y, Okamoto M. Tonsillar IgA1 as a possible source of hypoglycosylated IgA1 in the serum of IgA nephropathy patients. *Nephrol Dial Transplant* 2003; 18: 1108–14.
12. Linossier MT, Palle S, Berthoux F. Different glycosylation profile of serum IgA1 in IgA nephropathy according to the glomerular basement membrane thickness: normal versus thin. *Am J Kidney Dis* 2003; 41: 558–64.
13. van den Wall Bake AW, Daha MR, Radl J, Haaijman JJ, Van der Ark A, Valentijn RM, Van Es LA. The bone marrow as production site of the IgA deposited in the kidneys of patients with IgA nephropathy. *Clin Exp Immunol* 1988; 72: 321–5.
14. Harper SJ, Allen AC, Pringle JH, Feehally J. Increased dimeric IgA producing B cells in the bone marrow in IgA nephropathy determined by in situ hybridisation for J chain mRNA. *J Clin Pathol* 1996; 49: 38–42.
15. Béné MC, Faure G, Hurault de Ligny B, Kessler M, Duheille J. Immunoglobulin A nephropathy. Quantitative immunohistomorphometry of the tonsillar plasma cells evidences an inversion of the immunoglobulin A versus immunoglobulin G secreting cell balance. *J Clin Invest* 1983; 71: 1342–7.
16. Egido J, Blasco R, Lozano L, Sancho J, Garcia-Hoyo R. Immunological abnormalities in the tonsils of patients with IgA nephropathy: inversion in the ratio of IgA: IgG bearing lymphocytes and increased polymeric IgA synthesis. *Clin Exp Immunol* 1984; 57: 101–6.
17. Garcia-Hoyo R, Egido J, Lozano L, de Nicolas R, Hernando L. Disturbances of IgA immune regulation in lymphocytes from mucosae and peripheral blood in patients with IgA nephropathy. *Semin Nephrol* 1987; 7: 301–5.
18. Nagy J, Brandtzaeg P. Tonsillar distribution of IgA and IgG immunocytes and production of IgA subclasses and J chain in tonsillitis vary with the presence or absence of IgA nephropathy. *Scand J Immunol* 1988; 27: 393–9.
19. Béné MC, Hurault De Ligny B, Kessler M, Faure GC. Confirmation of tonsillar anomalies in IgA nephropathy: a multicenter study. *Nephron* 1991; 58: 425–8.
20. Harper SJ, Allen AC, Bene MC, Pringle JH, Faure G, Lauder I, Feehally J. Increased dimeric IgA-producing B cells in tonsils in IgA nephropathy determined by in situ hybridization for J chain mRNA. *Clin Exp Immunol* 1995; 101: 442–8.
21. Harper SJ, Pringle JH, Wicks AC, Hattersley J, Layward L, Allen A, Gillies A, Lauder I, Feehally J. Expression of J chain mRNA in duodenal IgA plasma cells in IgA nephropathy. *Kidney Int* 1994; 45: 836–44.
22. Brandtzaeg P, Halstensen TS, Kett K, Krajci P, Kvale D, Rognum TO, Scott H, Sollid LM. Immunobiology and immunopathology of human gut mucosa: humoral immunity and intraepithelial lymphocytes. *Gastroenterology* 1989; 97: 1562–84.
23. Béné MC, Faure G, Duheille J. IgA nephropathy: characterization of the polymeric nature of mesangial deposits by in vitro binding of free secretory component. *Clin Exp Immunol* 1982; 47: 527–34.
24. Conley ME, Cooper MD, Michael AF. Selective deposition of immunoglobulin A1 in immunoglobulin A nephropathy, anaphylactoid purpura nephritis, and systemic lupus erythematosus. *J Clin Invest* 1980; 66: 1432–6.
25. Tomino Y, Endoh M, Nomoto Y, Sakai H. Immunoglobulin A1 and IgA nephropathy. *N Engl J Med* 1981; 305: 1159–60.
26. Clarkson AR, Woodroffe AJ, Aarons I, Hiki Y, Hale G. IgA nephropathy. *Annu Rev Med* 1987; 38: 157–68.
27. Kennel-De March A, Béné MC, Hurault de Ligny B, Kessler M, Faure GC. Enhanced expression of CD31 and CD54 on tonsillar high endothelial venules in IgA nephropathy. *Clin Immunol Immunopathol* 1997; 84: 158–65.
28. Kennel-De March A, Béné MC, Renoult E, Kessler M, Faure GC. Low levels of spontaneously activated peripheral IgA-secreting cells in nontransplanted IgA nephropathy patients. *Am J Kidney Dis* 1997; 30: 64–70.
29. Kennel-de March A, Béné MC, Renoult E, Kessler M, Faure GC, Kolopp-Sarda MN. Enhanced expression of L-selectin on peripheral blood lymphocytes from patients with IgA nephropathy. *Clin Exp Immunol* 1999; 115: 542–6.
30. Tokuda M, Shimizu J, Sugiyama N, Kiryu T, Matsuoka K, Sasaki O, Fukuda K, Hatase O, Monden H. Direct evidence of the production of IgA by tonsillar lymphocytes and the binding of IgA to the glomerular mesangium of IgA nephropathy patients. *Acta Otolaryngol* 1996; Suppl. 523: 182–4.
31. Kodama S, Suzuki M, Arita M, Mogi G. Increase in tonsillar germinal centre B-1 cell numbers in IgA nephropathy (IgAN) patients and reduced susceptibility to Fas-mediated apoptosis. *Clin Exp Immunol* 2001; 123: 301–8.
32. Suzuki S, Fujieda S, Sunaga H, Sugimoto H, Yamamoto C, Kimura H, Abo T, Gejyo F. Immune response of tonsillar lymphocytes to *Haemophilus parainfluenzae* in patients with IgA nephropathy. *Clin Exp Immunol* 2000; 119: 328–32.
33. Vialtel P, Dechelette E, Hachache T, Colomb H, Cordonnier D, Roux O, Dumas G, Accoyer B, Junier Lavillauroy C, Charachon R. Rôle de l'amygdalectomie sur l'évolution des symptômes de la glomérulonéphrite à dépôts mésangiaux d'IgA. *Semin Urol Néphrol* 1981; 10: 173–85.

34. Tamura S, Masuda Y, Inokuchi I, Terasawa K, Sugiyama N. Effect of and indication for tonsillectomy in IgA nephropathy. *Acta Otolaryngol* 1993; Suppl. 508: 23–8.
35. Tomioka S, Miyoshi K, Tabata K, Hotta O, Taguma Y. Clinical study of chronic tonsillitis with IgA nephropathy treated by tonsillectomy. *Acta Otolaryngol* 1996; Suppl. 523: 175–7.
36. Xie Y, Nishi S, Ueno M, Imai N, Sakatsume M, Narita I, Suzuki Y, Akazawa K, Shimada H, Arakawa M, Gejyo F. The efficacy of tonsillectomy on long-term renal survival in patients with IgA nephropathy. *Kidney Int* 2003; 63: 1861–7.
37. Béné MC, Hurault de Ligny B, Kessler M, Foliguet B, Faure GC. Tonsils in IgA nephropathy. *Contrib Nephrol* 1993; 104: 153–61.

Address for correspondence:
Pr Marie C Béné
Lab Immunology, Faculté de Médecine
BP 184
FR-54500 Vandoeuvre les Nancy
France
Fax: +33 383 446 022
E-mail: Marie-Christine.Bene@medicine.uhp-nancy.fr

Copyright of Acta Oto-Laryngologica is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.

Copyright of *Acta Oto-Laryngologica* (Supplement) is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.